

Determination of Essential and Non-essential Metals Concentration in Garlic (*Allium sativum* L.) Bulb and Leaf Cultivated in Ambo Woreda, Ethiopia

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Abstract: In this study, the levels of twelve essential metals (Na, K, Ca, Mg, Fe, Zn, Mn, Cu, Mo, Co, Cr and Ni) and two non-essential metals (Pb and Cd) were determined in the bulb and leaf of garlic (*Allium sativum* L.) cultivated in Ambo Woreda, Ethiopia. Wet digestion method using a mixture of 5 ml of concentrated $\text{HNO}_3\text{:HClO}_4$ (4:1 v/v) was used for digestion of the samples. The determination processes were done by flame photometer for Na and K, EDTA titration method for Ca and Mg, and ICP-OES for the rest of the metals. The results obtained revealed that the concentrations of metals in the garlic bulb samples in mg/kg dry weight were in the range of: Na (217–366.7), K (9080–12060), Ca (1018–1286), Mg (802–992.6), Fe (63.44–91.24), Zn (31.17–35.39), Mn (5.27–7.51), Cu (4.21–7.16), Mo (1.06–2.08), Co (0.61–1.49), Ni (1.45–3.78), Cr (0.47–1.31), Pb (1.07–2.51) and Cd (0.10–0.16). The concentrations of metals in the garlic leaf samples in mg/kg dry weight were in the range of: Na (463–730), K (11370–12860), Ca (1209–1302), Mg (871–994), Fe (72.3–108), Zn (49.1–71.39), Mn (26.74–72.36), Cu (5.41–8.44), Mo (1.01–2.30), Co (1.17–4.96), Ni (2.17–3.54), Cr (1.20–2.17), Pb (1.87–2.84) and Cd (0.12–0.18). In addition, the results show that the levels of elements were higher in the leaves than the bulbs. In general, the levels of metals in the analyzed garlic bulb and leaf samples were found below the FAO/WHO maximum permissible limit; hence they are safe for human consumption and can be considered as a good source of essential nutrients.

Keywords: Essential Metals, Non-essential Metals, Garlic (*Allium sativum* L.), Wet Digestion, ICP-OES

1. Introduction

Garlic (*Allium sativum* L.), the second most important *Allium* crop next to the onion (*Allium cepa* L.), is cultivated worldwide and consumed by almost every culture as a popular condiment and green vegetable [1]. Garlic has been used throughout its history for both culinary and medicinal purposes [2]. Garlic (*Allium sativum* L.) is the most widely used bulb vegetable next to onion in Ethiopia. It contributes significant nutritional value to the human diet [3]. Garlic (*Allium sativum* L.) has a long folklore history as a treatment for cold, cough, asthma and is reported to strengthen the immune system. It has many medicinal effects such as lowering of blood cholesterol level, antiplatelet aggregation, antihelmantic, anti-inflammatory activity and inhibition of cholesterol synthesis. Garlic has long been known to have

antibacterial, antifungal, anticancer and antiviral properties [4, 5].

Pollution of foods by heavy metals is a worldwide phenomenon. Studies have revealed that fruits and leafy vegetables are vulnerable to heavy metal contamination from soil, wastewater and air pollution. The toxicity and consequent threat to human health by heavy metals such as cadmium, copper, lead, chromium, zinc, nickel, cobalt, arsenic and mercury are a function of concentration and bioaccumulation [6, 7]. The implications associated with metal contamination are of great concern, particularly in agricultural production systems due to their increasing trends in human foods and environment [8]. They are ubiquitous in the environment through various pathways, due to natural and anthropogenic activities [9]. Source of anthropogenic contamination include the addition of manures, sewage

sludge, fertilizers and pesticides to soils, several studies identifying the risks in relation to increased soil metal concentration and consequent plant uptake [10].

Heavy metals are non-biodegradable and bioaccumulate in living tissues through the food chain. This fact necessitates for frequent determination of heavy metals in vegetables and soil for the safety of consumers. Fruits, vegetables and other foods are among pathways by which heavy metals enter the human tissues leading to deterioration of health [7]. Excessive amount of heavy metals in food cause a number of diseases, especially cardiovascular, renal, neurological, and bone diseases [10, 11].

Elements such as nickel, zinc, iron, copper and magnesium etc. are essential because they are associated with enzyme systems and other biochemical processes in the body. In spite of their benefits, some trace elements cannot be regarded as essential to life. Lead, cadmium, arsenic, and mercury are toxic at very low concentrations and are termed non-essential. Cadmium and lead in any concentrated can caused kidney damage and toxicity symptoms include impaired kidney function, poor reproductive capacity, hypertension, tumors, etc. [12]. However, whether essential or not, when bioaccumulated, once they exceed the total body burden, disease conditions may arise [13].

Minerals are nutritionally important components in food, they are necessary for health, and are part of all aspect of cellular function. They are involved in structural components and also form an integral part of enzyme or protein structure. Minerals are essential for growth, development and maintenance of tissues and are also linked to the expression of genetic information, the effectiveness of immune system,

the prevention of cell damage. In general, minerals increase resistance to many chronic and some infectious diseases [14].

However, studies are mainly focused on the medicinal values of garlic; the study which was conducted on the nutritional values of garlic is scarce. Before the commencement of this work, there are no literature reports on the content of essential and non-essential metals in *Allium sativum* L. (garlic) cultivated in Ambo Woreda. So the main objective of this study was to determine the levels of essential metals (Na, K, Ca, Mg, Fe, Zn, Mn, Cu, Mo, Co, Ni and Cr) and non-essential metals (Pb and Cd) in garlic bulb and leaf samples and their comparative distribution in different locations of Ambo Woreda, West Shoa Zone of Oromia Regional State, Ethiopia for Na and K using flame photometer, Ca and Mg by EDTA titration method, and the rest of the metals using inductively coupled plasma-optical emission spectrometry.

2. Materials and Methods

2.1. Study Area

The study was conducted in 4 locations of Ambo Woreda (Awaro Qora, Gosu Qora, Qibafkuba and Elamu Goromti) Kebeles in West Shoa Zone of Oromia Region which is about 112 km West of Addis Ababa, the capital city of Ethiopia. Ambo is located between latitude 8°59'N and longitude 37°51'E with an elevation of 2101 meters above sea level, The temperature ranges from 15°C-29°C with average temperature of 22°C (Figure 1).

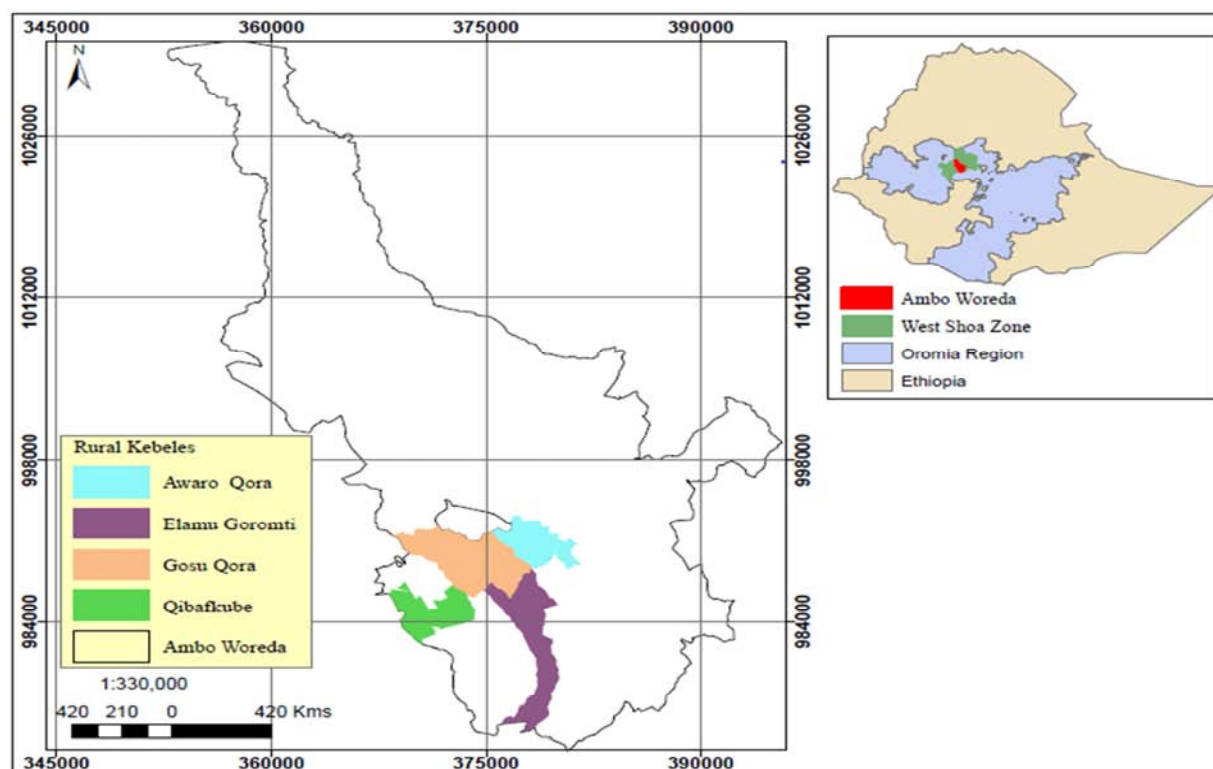


Figure 1. Map of the study area.

2.2. Chemicals and Reagents

All the chemicals used for this work were of analytical grades obtained from Uni-Chem® India. 65–68% nitric acid (HNO_3) and 70–72% perchloric acid (HClO_4) were used for sample digestion. 1000 mg/L of stock standard solution of each of the metals to be determined were used for the preparation of spiking and calibration standards. Double distilled water was used throughout the work.

2.3. Instrument and Apparatuses

Inductively coupled plasma-optical emission spectrometry (ICP-OES) model Agilent 720 was used for the determination of Fe, Zn, Mn, Cu, Mo, Co, Ni, Cr, Pb and Cd. Flame photometry model ELICO CL-378 was used for the determination of Na and K, while Ca and Mg were determined by EDTA titration method. Drying oven (Model DHG-9070A, Shanghai, China) was used for drying the garlic bulb and leaf samples. An analytical balance with an accuracy of ± 0.0001 g (Model AA-200DS, Deriver Instrument Company, Germany) was used for weighing the samples. Digestive Furnace (Model: KDN-20C China) was used for sample digestion. All flasks and glassware were washed with tap water using detergent, immersed in 10% (v/v) HNO_3 solution for 24 hours, and rinsed thoroughly with double distilled water.

2.4. Sample Collection

The garlic samples were collected in February, 2015 from the four agricultural Kebeles of Ambo Woreda (Awaro Qora, Gosu Qora, Qibafkuba and Elamu Goromti). From each sample site, garlic samples were collected from five different sub-sites (farm lands) to provide replicate samples of each site. These farmlands were chosen randomly. Five fresh garlic samples were collected from centers and corners of each sub-farm land. The five sub-samples were mixed together to form a composite sample that represent each sampling areas. Finally, four garlic bulk samples one from each stated areas were collected and put in clean plastic bags labeled and brought into the laboratory for further treatment.

2.5. Sample Treatment

The bulb and leaf parts of the garlic were separated with stainless steel Teflon knives, the bulbs were peeled and rinsed with tap water and then with distilled water to eliminate adsorbed dust particles. All the samples were cut into small sizes to facilitate drying of the pieces at the same rate and subsequently dried in the drying oven at 80°C for 48 hours to constant weight. The dried samples were ground into powder using mortar and pestle and then passed through a 0.5 mm mesh sieve. The samples were kept in plastic containers prior to analysis.

2.6. Preparation of Samples

0.5 g of each garlic bulb and leaf samples were taken in to

a digestive tubes containing a mixture of 5 ml of concentrated $\text{HNO}_3\text{:HClO}_4$ (4:1 v/v). The mixture was digested in a digestive furnace (Model: KDN-20C, China) by setting the temperature first 150°C for the first 1 hour, then increasing to 175°C for the remaining 2 hours. The digest was allowed to cool for 10 minutes without dismantling the condenser and further 10 minutes after removing the condenser. The digest was then diluted with 10 ml double distilled water and filtered through Whatman filter paper No. 41 in to 50 ml volumetric flask. The digestive flask further rinsed with 10 ml double distilled water and added to the filtrate, and the flask containing the filtrate was made up to the mark with double distilled water. Each sample was digested in triplicate and transferred into clean and dry plastic bottles, labeled and stored in refrigerator at 4°C until analysis by ICP-OES. The blank solutions were undergoing the same digestion procedure as that of the samples.

2.7. Preparation of Calibration Standards and Spiking Standards

For calibration of the instruments, a series of five standard solutions were prepared by appropriate dilutions from 1000 ppm stock standard solution of the metals to be analyzed into 100 ml volumetric flasks. The prepared metal concentrations include: 0.2, 0.5, 1, 1.5 and 2 ppm of Fe, Zn, Mn, Cu, Mo, Co, Ni, Cr, Pb and Cd, and 1, 2, 4, 8, 16 ppm of Na and K.

For the spiking processes, a mixture of standard solution containing 4 mg/L of each Na and K; 1mg/L of Fe, Zn, Mn, Cu, Mo, Co, Ni, Cr, Pb and Cd was prepared by serial dilution from 1000 mg/L stock standard solution in to 100 ml volumetric flask and diluting to the mark with double distilled water.

2.8. Method Validation

The proposed method was validated by evaluating different parameters such as linearity, matrix effect, limit of detection (LOD), limit of quantitation (LOQ), accuracy (in terms of recovery) and precision (in terms of repeatability) [15].

2.8.1. Accuracy and Precision

The accuracy and precision of the proposed procedure were evaluated by the analysis of matrix spike samples and laboratory control samples. Accuracy was evaluated through recovery studies of sample spikes. Precision was evaluated regarding repeatability by estimating the relative standard deviation (RSD) of the recovery percentage for each spiked level.

In this study, the recovery test was done by spiking a suitable known quantity of metal standard solution into a test portion of the sample. For doing so, each sample was spiked in triplicates at near mid-range calibration concentration (4.0 mg/L of each of Na and K; 1.0 mg/L of Ca, Mg, Fe, Zn, Mn, Cu, Mo, Co, Ni, Cr, Pb and Cd). The spiked and non-spiked samples were digested and analyzed using the same analytical procedure as the garlic sample. The percent

recoveries of the analyte were calculated by using equation 1 [16, 17].

$$\% \text{ Recovery} = \frac{\text{spiked result} - \text{unspiked result}}{\text{added amount}} \times 100 \quad (1)$$

Where, conc. = concentration of metal of interest.

The relative standard deviation for replicate analyses of the same sample was obtained as dividing the standard deviation by the mean value of the analytical data according to the following equation [16].

$$\% \text{ RSD} = \frac{S}{\bar{X}} \quad (2)$$

Where, S = standard deviation and \bar{X} the mean of the replicate analysis.

2.8.2. Limit of Detection

The limit of detection (LOD) is taken as the lowest concentration of an analyte in a sample that can be detected, but not necessarily quantified, under the stated conditions of the test. LOD is calculated as:

$$\text{LOD} = 3S_a/b \quad (3)$$

Where S_a is the standard deviation of the response; can be obtained by standard deviation of blank, response residual standard deviation of the regression line, or standard deviation of the y-intercept of the regression line and b is the slope of the calibration curve [18, 19]. In this study, the LOD was obtained from triplicate analysis of reagents blanks which were digested in the same digestion procedure as the actual samples. The LOD for each analyte was calculated and the results are presented in Table 1.

2.8.3. Limit of Quantitation

The limit of quantitation (LOQ) is the lowest concentration of an analyte in a sample that can be quantitatively determined with acceptable precision and accuracy under the stated conditions of test [18, 19]. LOQ is calculated as:

$$\text{LOQ} = 10S_a/b \quad (4)$$

Where S_a is the standard deviation of the response and b is the slope of the calibration curve [18]. In this study, LOQ was obtained from triplicate analysis of reagents blanks which were digested in the same digestion procedure as the actual samples. The LOQ for each analyte was calculated and the results are indicated in Table 1.

2.8.4. Contamination Control

Many measurement processes are prone to contamination, which can occur at any point in the sampling, sample preparation, instrument or analysis [20]. Therefore, to control this contamination effects the following controlling mechanisms are performed.

Calibration Blank: To measure the amount of the analytical signal that arises from the solvents and reagents; a calibration blank of 2% HNO_3 was prepared and run together with the standards for creating zero concentration point of the

calibration graph. This helps to establish the baseline of an instrument.

Method Blank: Method blank is an analyte-free sample carried through the analysis using the same reagents, glassware and instrumentation. Method blanks are used to identify and correct systematic errors due to impurities in the reagents, contamination in the glassware and instrumentation [21].

In this study, 0.5 g sucrose was used as matrix for the garlic bulb and leaf samples. The blanks which were prepared from sucrose were treated exactly like the sample including exposure to all glassware, digestion media, apparatus, solvents and reagents that are used with other samples but with no added sample.

Laboratory Control Sample: The laboratory control sample (LCS) was analyzed in an identical manner as a sample and the results were used to assess accuracy and precision of the analytical methodology. Results within $\pm 10\%$ of the true value are accepted. In this work, 0.5 g sucrose were spiked with 4.0 mg/L of each of Na and K; 1.0 mg/L of Ca, Mg, Fe, Zn, Mn, Cu, Mo, Co, Ni, Cr, Pb and Cd. The spiked samples were digested like the garlic samples including exposure to all glassware, digestion media, apparatus, solvents and reagents that are used with the garlic samples. The percent LCS recoveries for each metal were calculated using the following equation [22].

$$\% R = \frac{\text{LCS} - \text{MB}}{S} \times 100 \quad (5)$$

Where: % R = percent recovery, LCS = Laboratory Control Sample Results, S = amount of spike added and MB = results of the method blank

2.9. Elemental Analyses of Samples

The digested garlic bulb and leaf samples were analyzed for Ca and Mg by EDTA titration method; Na and K using flame photometer (Model: ELICO CL-378) and Fe, Zn, Mn, Cu, Mo, Co, Ni, Cr, Pb and Cd using Agilent 720 Inductively Coupled Plasma Optical Emission Spectrometry (ICP-OES) after calibrating the instruments using calibration blank and five series of working standard solutions of each metal to be analyzed. Final concentration of element in the samples was calculated as:

$$\text{Concentration (mg/kg)} = \frac{\text{Concentration (mg/L)} \times V}{W} \quad (6)$$

Where: V is the final volume of the digested solution (50 ml) and W is the weight of the sample (0.5 g).

2.10. Statistical Analysis

All analyses were carried out in triplicates and the data were presented as means \pm standard deviations. One-way analysis of variance (ANOVA) at $P < 0.05$ was used to determine statistically significant differences in the mean concentrations of metals among groups of garlic bulb and leaf samples. Pearson's correlation analysis was also applied

to test the correlation between metals in garlic bulb and leaf samples. A probability level of $P < 0.05$ was considered statistically significant. All statistical analyses were done by SPSS version 16.0 software for windows.

3. Results and Discussion

3.1. Method Validation Results

3.1.1. Calibration Curves, Limit of Detection and Limit of Quantitation

Table 1 shows the wavelength used for the ICP-OES elemental analysis, the calibration curve equation, the correlation coefficients, the limits of detection (LOD), and

limits of quantitation (LOQ) of the trace elements analyzed in the garlic bulb and leaf samples.

For all analytes, the analytical curves showed correlation coefficients (R) values higher than 0.999, indicating a good linear correlation between the analytical signal and the analyte concentration.

From Table 1, the limit of detection (LOD) values for all the metals analyzed ranged from 0.1–0.8 $\mu\text{g/g}$ and the limit of quantitation (LOQ) values for all the metals analyzed ranged from 0.3–2.5 $\mu\text{g/g}$. The LOD and LOQ method obtained were low enough to detect the presence of metals of interest at trace levels in both samples.

Table 1. Wavelength of detection, calibration curve equation, correlation coefficient (R) of the calibration curves, limit of detection (LOD) and limit of quantitation (LOQ) obtained for each element.

Element	Wavelength (nm)	Calibration equation	R	LOD ($\mu\text{g/g}$)	LOQ ($\mu\text{g/g}$)
Fe	238.2	$Y = 2068.4x + 18.7$	0.9998	0.8	2.5
Zn	206.2	$Y = 4226.9x + 26.3$	0.9999	0.3	1.2
Mn	294.9	$Y = 75560.8x + 76.3$	0.9999	0.3	1.0
Cu	324.7	$Y = 22876.2x + 180.1$	0.9999	0.3	1.2
Mo	201.5	$Y = 2295.5x + 24.8$	0.9998	0.3	1.2
Co	236.4	$Y = 2348.0x + 21.8$	0.9999	0.6	1.9
Ni	221.6	$Y = 2068.4x + 18.7$	0.9999	0.6	1.9
Cr	206.5	$Y = 4570.1x + 60.3$	0.9999	0.5	1.6
Pb	283.3	$Y = 1050.3x + 23.4$	0.9999	0.3	1.2
Cd	214.4	$Y = 1617.2x + 17.8$	0.9997	0.1	0.3

3.1.2. Precision and Accuracy

The precision and accuracy of the proposed method were evaluated by means of matrix spike recovery tests. The recovery values of triplicate analysis of the matrix spike garlic bulb and leaf samples were calculated using equation 1 and the RSD values were calculated using equation 2 and the results are presented in Table 2 and 3 respectively.

Table 2. Recovery and precision test results of metals for garlic bulb matrix spike sample.

Metals	Conc. in sample ($\mu\text{g/g}$)	Amount added ($\mu\text{g/g}$)	Conc. in spiked sample ($\mu\text{g/g}$)	Recovery (%)	RSD (%)
Na	360 ± 8.16	400	734.2 ± 2.10	93.55 ± 3.21	3.43
K	9080 ± 143.3	400	9488 ± 4.20	102.00 ± 2.74	2.69
Ca	1211 ± 47.28	100	1301.3 ± 7.34	90.30 ± 7.38	8.17
Mg	876 ± 57.45	100	967.4 ± 9.16	91.40 ± 9.12	9.98
Fe	89.56 ± 0.09	100	180.7 ± 4.72	91.14 ± 4.63	5.08
Zn	31.68 ± 0.05	100	124.2 ± 1.15	92.52 ± 2.50	2.70
Mn	5.38 ± 0.05	100	104.31 ± 5.41	98.93 ± 1.94	1.96
Cu	4.21 ± 0.16	100	99.8 ± 2.82	95.59 ± 2.71	2.84
Mo	2.08 ± 0.18	100	95.3 ± 1.17	93.20 ± 4.11	4.41
Co	0.85 ± 0.09	100	92.2 ± 3.54	91.35 ± 3.60	3.94
Ni	1.45 ± 0.02	100	94.0 ± 5.68	92.55 ± 7.09	7.66
Cr	0.54 ± 0.06	100	90.96 ± 1.63	90.42 ± 2.16	2.39
Pb	2.02 ± 0.56	100	93.27 ± 1.45	91.25 ± 5.08	5.57
Cd	0.16 ± 0.05	100	93.78 ± 2.71	93.62 ± 3.24	3.46

Table 3. Recovery and precision test results of metals for garlic leaf matrix spike sample.

Metals	Conc. in sample ($\mu\text{g/g}$)	Amount added ($\mu\text{g/g}$)	Conc. in spiked sample ($\mu\text{g/g}$)	Recovery (%)	RSD (%)
Na	730 ± 8.16	400	1106.4 ± 3.41	94.10 ± 4.26	4.53
K	12520 ± 165.1	400	12915.2 ± 5.13	98.80 ± 3.18	3.22
Ca	1250 ± 81.32	100	1342.7 ± 2.45	92.70 ± 6.27	6.76
Mg	942 ± 65.21	100	1033.4 ± 7.29	91.40 ± 1.54	1.68
Fe	94.20 ± 1.89	100	185.51 ± 4.72	91.31 ± 5.51	6.03
Zn	49.10 ± 0.26	100	141.2 ± 2.68	92.10 ± 3.47	3.77
Mn	26.74 ± 0.09	100	118.0 ± 2.43	91.26 ± 4.55	4.99
Cu	8.44 ± 2.13	100	105.3 ± 3.16	96.86 ± 2.16	2.23

Metals	Conc. in sample ($\mu\text{g/g}$)	Amount added ($\mu\text{g/g}$)	Conc. in spiked sample ($\mu\text{g/g}$)	Recovery (%)	RSD (%)
Mo	2.27 ± 0.07	100	97.1 ± 5.62	94.83 ± 2.53	2.67
Co	2.09 ± 0.14	100	94.35 ± 4.37	92.26 ± 5.14	5.57
Ni	2.17 ± 0.01	100	92.7 ± 7.21	90.53 ± 8.90	9.83
Cr	1.51 ± 0.23	100	95.3 ± 3.72	93.79 ± 6.24	6.65
Pb	2.48 ± 0.41	100	93.54 ± 1.45	91.06 ± 3.79	4.16
Cd	0.17 ± 0.06	100	93.07 ± 3.72	92.90 ± 2.18	2.35

As it can be seen in Table 2, the percentage recovery of the metal analysis in the garlic bulb samples ranged between 90.30–102.00% and the RSD values ranged between 1.96–9.98%. From Table 3, percentage recovery of the metal analysis in the garlic leaf ranged between 90.53–98.80% and the RSD values ranged between 1.68–9.83%. The matrix spike recovery obtained in this study falls within the acceptable range of 90–110% for a good recovery study. The high percentage recovery obtained from the study validates the accuracy of the method and the reliability of the levels of metal concentration in this study. The RSD values of the samples were $< 10\%$, indicating that the proposed method was precise.

3.1.3. Calibration Control

Analysis of metal standard solution of mid-point calibration curves after every 10 sample and at the end of sample run shows that each analyte falls $\pm 10\%$ of the expected value. This indicates that the sample analysis is within the control limits.

3.1.4. Contamination Control

Method blanks were run to identify and correct systematic errors due to impurities in the reagents and contamination in the glassware and instrumentation. The analysis results revealed that there were no readings above the method detection limits of the metals. Hence, it can be concluded that the analytical method was free of overall laboratory contamination.

3.1.5. Laboratory Control Sample Results

Table 4. Recovery and precision test results for the laboratory control samples.

Metals	Amount added ($\mu\text{g/g}$)	Conc. in spiked sample ($\mu\text{g/g}$)	Recovery (%)	RSD (%)
Na	400	392.4 ± 1.23	98.08 ± 2.47	2.52
K	400	374.48 ± 2.15	93.56 ± 5.04	5.39
Ca	100	90.41 ± 2.37	90.34 ± 6.21	6.87
Mg	100	93.41 ± 1.86	93.36 ± 3.85	4.12
Fe	100	94.75 ± 5.18	94.68 ± 1.97	2.08
Zn	100	101.25 ± 7.15	101.19 ± 3.16	3.12
Mn	100	90.63 ± 8.65	90.56 ± 8.94	9.87
Cu	100	99.82 ± 7.23	99.65 ± 3.15	3.16
Mo	100	90.85 ± 3.16	90.79 ± 5.42	5.97
Co	100	91.27 ± 4.20	91.20 ± 1.94	2.13
Ni	100	92.83 ± 3.17	92.75 ± 8.43	9.09
Cr	100	91.66 ± 1.96	91.58 ± 2.16	2.36
Pb	100	90.79 ± 2.47	90.72 ± 3.43	3.78
Cd	100	92.71 ± 1.58	92.65 ± 2.98	3.22

Laboratory control sample recoveries and relative standard deviations were calculated for the triplicate analysis of each analyte using equation 5 and 2 respectively. The results are summarized in Table 4. As can be seen from the table, the percent recovery values of laboratory control sample (LCS) results lied in the range of 90.34–101.19% and the RSD values ranged from 2.08–9.87%. The percent recovery obtained in this study falls within the normal acceptable range of 90–110% for a good LCS recovery study and $\leq 10\%$ for RSD. These results showed that the analytical method possesses the required precision and accuracy.

3.2. Results of the Determination of Essential and Non-Essential Metals

3.2.1. Levels of Metals in Garlic (*Allium sativum* L.) Bulb Samples

In the present study, the mean concentrations of the studied essential and non-essential metals in garlic (*Allium sativum* L.) bulb are given in Table 5.

As can be seen from Table 5, the sodium content in the garlic bulb samples ranged from 217 to 366.7 mg/kg. The lowest concentration of sodium (217 mg/kg) was found in garlic bulb collected from Qibafkuba site and highest concentration of sodium (366.7 mg/kg) was found in garlic bulb collected from Gosu Qora site. Whereas, Elamu Goromti and Awaro Qora garlic bulb contains 343 and 360 mg/kg respectively. The levels indicated that Na is the least accumulated metal by garlic bulb among the four macroelements determined in all the samples.

The concentrations of potassium in the garlic bulb samples were found higher than all the metals analyzed. Mean potassium concentration ranged from 9080 mg/kg to 12060 mg/kg. The lowest and highest concentration of K was found in Awaro Qora (9080 mg/kg) and Elamu Goromti (12060 mg/kg) sites respectively. The amount of potassium in Gosu Qora and Qibafkuba garlic samples were 10258 mg/kg and 11580 mg/kg respectively (Table 5). However, there was significant difference ($p < 0.05$) in the content of potassium between the sampling sites.

Calcium was the second most accumulated essential metal next to potassium in the garlic bulb. The average concentration of calcium (Table 5) in the garlic bulb samples ranged from 1018 mg/kg in Elamu Goromti garlic bulb to 1286 mg/kg in Gosu Qora garlic bulb. The calcium content of Qibafkuba and Awaro Qora garlic bulb were 1079 and 1211 mg/kg respectively.

The level of magnesium in garlic bulbs ranged from 802 to 992.6 mg/kg. However, there was significant difference ($p <$

0.05) in the content of Mg between the sampling sites. The lowest concentration (802 mg/kg) being in garlic collected from Qibafkuba site and the highest concentration (992.6 mg/kg) in garlic collected from Gosu Qora site. The amount of Mg in Elamu Goromti and Awaro Qora garlic bulb were 854 mg/kg and 876 mg/kg respectively.

The results in Table 5 reveal that the concentrations of iron in the garlic bulb samples were 63.44 mg/kg in Elamu Goromti garlic bulb, 85.82 mg/kg in Qibafkuba garlic bulb, 89.56 mg/kg in Awaro Qora garlic bulb and 91.24 mg/kg in Gosu Qora garlic bulb. The results found were lower than the FAO/WHO maximum permissible limit for medicinal plant that is 425 mg/kg [23]. One-way ANOVA test showed that there was significant difference ($P < 0.05$) among the mean concentrations of iron in the garlic bulb.

Zinc level in garlic bulb was in the range of 31.17–35.39 mg/kg as shown in Table 5. The lowest zinc content was obtained in garlic bulbs collected from Gosu Qora site and the highest in garlic bulb collected from Qibafkuba site. However, there was no significant variation ($p > 0.05$) in the content of Zn between the sampling sites. The FAO/WHO recommended limit of zinc in medicinal plant is 100 mg/kg [23].

As evident from Table 5, the manganese contents of garlic bulb in all the sampling sites were almost similar except Gosu site. The obtained results are 5.27 mg/kg for Qibafkuba, 5.38 mg/kg for Awaro Qora, 5.49 mg/kg for Elamu Goromti, and 7.51 mg/kg for Gosu Qora garlic bulb. However, there was no significant difference ($p > 0.05$) in the content of Mn between the sampling sites except Gosu Qora site. The level of Mn found in this study was lower than the FAO/WHO (2001) [23] recommended limit of 500 mg/kg.

The concentration of copper in garlic bulb samples ranged between 4.21 to 7.16 mg/kg (Table 5). The lowest concentration of copper (Cu) was found in garlic bulb sample collected from Awaro Qora site while the highest concentration was found in Qibafkuba site. The content of Cu reported in this study was generally lower than the permissible levels by FAO/WHO (2001) in vegetables [23]. One-way ANOVA test showed that there was significant difference ($P < 0.05$) among the mean concentrations of Cu in the garlic bulb samples.

Table 5 shows the molybdenum level in garlic bulb was in the range of 1.06–2.08 mg/kg. The lowest molybdenum content was obtained in garlic bulb collected from Elamu Goromti site (1.06 mg/kg) and the highest in garlic bulb collected from Awaro Qora site (2.08 mg/kg). The concentration of molybdenum in Gosu Qora and Qibafkuba garlic bulbs were 1.33 and 1.47 mg/kg respectively. However, there was no significant variation ($p > 0.05$) in the level of molybdenum between the sampling sites.

The results of cobalt concentration in garlic bulb samples were 0.61, 0.85, 0.99 and 1.49 mg/kg in sample sites of Elamu Goromti, Awaro Qora, Qibafkuba and Gosu Qora,

respectively (Table 5). The level of cobalt found in this study was lower than the FAO/WHO (2001) [23] recommended limit that is 50 mg/kg. One-way ANOVA test showed that there was significant difference ($P < 0.05$) among the mean concentrations of Co in the garlic bulb samples.

As can be shown in Table 5, the average concentrations of nickel in garlic bulb grown in Awaro Qora, Gosu Qora, Qibafkuba and Elamu Goromti were 1.45, 2.45, 3.78 and 3.69 mg/kg, respectively. One-way ANOVA test showed that there was significant difference ($P < 0.05$) among the mean concentrations of nickel in the garlic bulb samples.

From Table 5, the mean concentrations of chromium in the garlic bulb samples were 0.54 mg/kg in Awaro Qora garlic bulb, 1.31 mg/kg in Gosu Qora garlic bulb, 0.81 mg/kg in Qibafkuba garlic bulb, and 0.47 mg/kg in Elamu Goromti garlic bulb. The concentrations of Cr obtained in this study were lower than the FAO/WHO (2001) [23] recommended maximum limit for plant that is 2.3 mg/kg. One-way ANOVA test showed that there was significant difference ($P < 0.05$) among the mean concentrations of Cr in the garlic bulb samples.

In the studied garlic bulb samples, the results of lead concentrations were 2.02, 2.51, 1.75 and 1.07 mg/kg in sample sites of Awaro Qora, Gosu Qora, Qibafkuba and Elamu Goromti, respectively (Table 5). The relatively high levels of lead might have resulted from accumulation of lead through air pollution and from some pesticides, such as lead arsenates applied during cultivation. The WHO (1998) [24] recommended maximum limit of Pb for medicinal plant is 10 mg/kg. The levels of lead found in this study were lower than this maximum permissible limit.

As can be seen from Table 5, the mean concentrations of cadmium in the garlic bulb samples were 0.16 mg/kg in Awaro Qora garlic bulb, 0.12 mg/kg in Gosu Qora garlic bulb, 0.10 mg/kg in Qibafkuba garlic bulb, and 0.11 mg/kg in Elamu Goromti garlic bulb. The high level of cadmium might be due to the use of cadmium-containing phosphate fertilizers and contamination from cadmium-containing dusts. The concentrations of cadmium obtained in this study were lower than the WHO (1998) [24] recommended maximum limit for medicinal plant that is 0.3 mg/kg.

In this study, we can observe that Na, Ca, Mg, Fe, Mn, Co, Cr and Pb were found to be present in highest concentration in the garlic bulb sample taken from Gosu Qora site. In the same manner, Zn and Cu were found to be highest in Qibafkuba garlic bulb. K and Ni were highest in Elamu Goromti garlic bulb. The highest value of Mo and Cd were found in Awaro Qora garlic bulb.

The result of the present study showed a high level of macro elements accumulation in the garlic bulb. In general, the mean concentrations of metals in garlic (*Allium sativum*) bulb collected from all sampling site decreased in the order of: $K > Ca > Mg > Na > Fe > Zn > Mn > Cu > Ni > Pb > Mo > Co > Cr > Cd$.

Table 5. Mean concentrations of metals (mg/kg dry weight) of garlic bulb samples.

mean \pm sd, n = 3.					
Sample Sites					Max. safe
Metals	Awaro Qora	Gosu Qora	Qibafkuba	Elamu Goromti	Limit in
	garlic bulb	garlic bulb	garlic bulb	garlic bulb	Plant (mg/kg)
Na	360 \pm 8.16	366.7 \pm 4.71	217 \pm 36.80	343 \pm 23.57	NA
K	9080 \pm 143.3	10258 \pm 125.6	11580 \pm 158	12060 \pm 122.3	NA
Ca	1211 \pm 47.28	1286 \pm 94.12	1079 \pm 78.29	1018 \pm 67.73	NA
Mg	876 \pm 57.45	992.6 \pm 85.34	802 \pm 42.81	854 \pm 38.56	NA
Fe	89.56 \pm 0.09	91.24 \pm 0.18	85.82 \pm 0.58	63.44 \pm 0.53	425 ^a
Zn	31.68 \pm 0.05	31.17 \pm 0.28	35.39 \pm 0.98	34.39 \pm 0.06	100 ^a
Mn	5.38 \pm 0.05	7.51 \pm 0.13	5.27 \pm 0.04	5.49 \pm 0.06	500 ^a
Cu	4.21 \pm 0.16	5.16 \pm 0.22	7.16 \pm 0.25	5.01 \pm 0.18	73 ^a
Mo	2.08 \pm 0.18	1.33 \pm 0.19	1.47 \pm 0.27	1.06 \pm 0.35	NA
Co	0.85 \pm 0.09	1.49 \pm 0.15	0.99 \pm 0.09	0.61 \pm 0.07	50 ^a
Ni	1.45 \pm 0.02	2.45 \pm 0.43	3.78 \pm 0.29	3.69 \pm 0.41	67 ^a
Cr	0.54 \pm 0.06	1.31 \pm 0.21	0.81 \pm 0.17	0.47 \pm 0.04	2.3 ^a
Pb	2.02 \pm 0.56	2.51 \pm 0.13	1.75 \pm 0.11	1.07 \pm 0.59	10 ^b
Cd	0.16 \pm 0.05	0.12 \pm 0.04	0.10 \pm 0.02	0.11 \pm 0.02	0.3 ^b

Key: ^aSource: [23], ^bSource: [24], NA= not available, sd = standard deviation.

3.2.2. Levels of Metals in Garlic (*Allium sativum* L.) Leaf Samples

In the present study, the mean concentrations of the studied essential and non-essential metals in garlic (*Allium sativum* L.) leaf are given in Table 6.

As can be seen from Table 6, the sodium content in the garlic leaf samples ranged from 463 to 730 mg/kg. The lowest concentration of Na (433 mg/kg) was found in garlic leaf collected from Qibafkuba site and highest concentration of Na (730 mg/kg) was found in garlic leaf collected from Awaro Qora. Whereas, Gosu Qora and Elamu Goromti garlic leaf contains 693 and 503 mg/kg respectively. The result indicated that Na is the least accumulated metal by garlic leaf among the four macro elements analyzed.

The concentration of potassium in the garlic leaf samples were found higher than all the metals analyzed. Mean potassium concentration ranged from 11370 mg/kg to 12860 mg/kg. However, there was significant difference ($p < 0.05$) in the content of K between the sampling sites. The lowest and highest concentration of potassium was found in Gosu Qora (11370 mg/kg) and Elamu Goromti (12860 mg/kg) sites. The amount of K in Awaro Qora and Qibafkuba garlic leaf were 12520 mg/kg and 11926 mg/kg respectively (Table 6).

Calcium was the second most accumulated essential metal next to potassium in garlic leaf samples. The average concentration of calcium in the garlic leaf samples ranged from 1209 mg/kg in Qibafkuba garlic leaf to 1302 mg/kg in Gosu Qora garlic leaf. The calcium content of Awaro Qora and Elamu Goromti garlic leaf were 1250 and 1264 mg/kg respectively (Table 6).

The level of magnesium in garlic leaf ranged from 871 to 994 mg/kg. However, there was significant difference ($p < 0.05$) in the content of magnesium between the sampling sites. The lowest concentration (871 mg/kg) being in garlic leaf collected from Qibafkuba site and the highest concentration (994 mg/kg) in garlic leaf collected from Gosu

Qora sampling site. The amount of magnesium in Elamu Goromti and Awaro Qora garlic leaf were 894 mg/kg and 942 mg/kg respectively (Table 6).

The results in Table 6 reveal that the concentrations of iron in the garlic leaf samples were 72.30 mg/kg in Elamu Goromti garlic leaf, 86.00 mg/kg in Qibafkuba garlic leaf, 94.20 mg/kg in Awaro Qora garlic leaf and 108.00 mg/kg in Gosu Qora garlic leaf. The results found were lower than the FAO/WHO maximum permissible limit for medicinal plant that is 425 mg/kg [23]. ANOVA test showed that there was significant difference ($P < 0.05$) among the mean concentrations of Fe in the garlic leaf samples.

Zinc level in garlic leaf was in the range of 49.1–71.39 mg/kg as shown in Table 6. The lowest Zn content was obtained in garlic leaf collected from Awaro Qora site and the highest in garlic leaf collected from Elamu Goromti site. However, there was significant variation ($p < 0.05$) in the content of Zn between all the sampling sites. The value of Zn obtained in this study was less than the FAO/WHO recommended limit of Zn in medicinal plant that is 100 mg/kg [23].

As evident from Table 6, the manganese contents of garlic leaf obtained were 26.74 mg/kg for Awaro Qora, 33.78 mg/kg for Gosu Qora, 58.67 mg/kg for Qibafkuba, and 72.36 mg/kg for Elamu Goromti garlic leaf samples. However, there was significant variation ($p < 0.05$) in the content of Mn between the sampling sites. The level of Mn found in this study was lower than the FAO/WHO (2001) recommended limit [23].

The concentration of copper (Table 6) in garlic leaf samples ranged between 5.41 to 8.44 mg/kg. The lowest concentration of Cu was found in garlic sample collected from Gosu Qora site while the highest concentration was found in Awaro Qora site. The content of Cu reported in this study was generally lower than the permissible levels by FAO/WHO (2001) [23] in vegetables. One-way ANOVA test

showed that there was significant difference ($P < 0.05$) among the mean concentrations of copper in the garlic leaf samples collected from all the sampling sites.

From Table 6, the concentration of molybdenum level in garlic leaf was in the range of 0.84–2.30 mg/kg. The lowest Mo content was obtained in garlic leaf collected from Qibafkuba site (0.84 mg/kg) and the highest in garlic leaf collected from Gosu Qora site (2.30 mg/kg). The concentration of Awaro Qora and Elamu Goromti garlic leafs were 2.27 and 1.01 mg/kg respectively. ANOVA test showed that there was significant difference ($P < 0.05$) among the mean concentrations of Mo in the garlic leaf samples.

The results of cobalt (Co) concentrations were 1.17, 2.09, 3.09 and 4.96 mg/kg in sample sites of Elamu Goromti, Awaro Qora, Qibafkuba and Gosu Qora, respectively (Table 6). The level of cobalt found in this study was lower than the FAO/WHO (2001) [23] recommended limit of 50 mg/kg. ANOVA test showed that there was significant difference ($P < 0.05$) among the mean concentrations of Co in the garlic leaf samples.

The average concentrations of nickel in garlic leaf in Awaro Qora, Gosu Qora, Qibafkuba and Elamu Goromti were 2.17, 3.54, 3.51 and 2.54 mg/kg, respectively (Table 6). One-way ANOVA test showed that there was significant difference ($P < 0.05$) among the mean concentrations of nickel in the garlic leaf samples.

From Table 6, the mean concentrations of Chromium (Cr) in the garlic samples were 1.51 mg/kg in Awaro Qora garlic leaf, 2.17 mg/kg in Gosu Qora garlic leaf, 1.32 mg/kg in Qibafkuba garlic leaf, and 1.20 mg/kg in Elamu Goromti garlic leaf. However, there was no significant variation ($p > 0.05$) in the content of Cr between the sampling sites. The concentrations of Cr obtained in this study were lower than the FAO/WHO (2001) [23] recommended maximum limit for

plant that is 2.3 mg/kg.

In the studied garlic leaf samples, the results of lead concentrations were 2.48, 1.87, 2.69 and 2.84 mg/kg in sample sites of Awaro Qora, Gosu Qora, Qibafkuba and Elamu Goromti, respectively (Table 6). However, there was no significant variation ($p > 0.05$) in the content of Pb between the sampling sites. The relatively high levels of Pb might have resulted from accumulation of Pb through air pollution and from some pesticides, such as lead arsenates applied during cultivation. The levels of Pb found in this study were lower than the WHO (1998) [24] recommended maximum limit for medicinal plant that is 10 mg/kg.

From Table 6, the mean concentrations of cadmium (Cd) in the garlic leaf in Awaro Qora, Gosu Qora, Qibafkuba and Elamu Goromti were 0.17, 0.18, 0.14 and 0.12 mg/kg, respectively. The relatively high level of Cd might be due to the use of cadmium-containing phosphate fertilizers and contamination from cadmium-containing dusts. The concentrations of cadmium obtained in this study were lower than the WHO (1998) [24] recommended maximum limit for medicinal plant that is 0.3 mg/kg.

In this study, we can observe that Ca, Mg, Fe, Mo, Co, Ni, Cr and Cd were found to be present in highest concentration in the garlic leaf sample taken from Gosu Qora site. In the same manner, Na and Cu were found to be highest in Awaro Qora garlic leaf. The highest value of K, Zn, Mn and Pb was found in Elamu Goromti garlic leaf sample.

The result of the present study showed a high level of macroelements accumulation in the garlic leaf. In addition, the results show that the levels of elements were higher in the leaves than the bulbs. In general, the mean concentrations of metals in garlic leaf collected from all sampling site decreased in the order of: $K > Ca > Mg > Na > Fe > Zn > Mn > Cu > Ni > Co > Pb > Mo > Cr > Cd$.

Table 6. Mean concentrations of metals (mg/kg dry weight) of garlic leaf samples.

mean \pm sd, n = 3.					
Sample Sites					Max. safe
Metals	Awaro Qora	Gosu Qora	Qibafkuba	Elamu Goromti	Limit in Plant
	garlic leaf	garlic leaf	garlic leaf	garlic leaf	(mg/kg)
Na	730 \pm 8.16	693 \pm 33.99	463 \pm 0.69	503 \pm 52.49	NA
K	12520 \pm 165	11370 \pm 111	11926 \pm 277	12860 \pm 257.8	NA
Ca	1250 \pm 81.32	1302 \pm 76.34	1209 \pm 78.5	1264 \pm 79.86	NA
Mg	942 \pm 65.21	994 \pm 78.53	871 \pm 67.43	894 \pm 65.28	NA
Fe	94.20 \pm 1.89	108 \pm 3.12	86 \pm 1.17	72.30 \pm 0.81	425 ^a
Zn	49.10 \pm 0.26	56.87 \pm 1.29	68.14 \pm 1.59	71.39 \pm 0.62	100 ^a
Mn	26.74 \pm 0.09	33.78 \pm 0.26	58.67 \pm 0.43	72.36 \pm 0.74	500 ^a
Cu	8.44 \pm 2.13	5.41 \pm 0.18	7.18 \pm 0.41	6.33 \pm 0.33	73 ^a
Mo	2.27 \pm 0.07	2.30 \pm 0.49	0.84 \pm 0.05	1.01 \pm 0.05	NA
Co	2.09 \pm 0.14	4.96 \pm 0.84	3.09 \pm 0.21	1.17 \pm 0.10	50 ^a
Ni	2.17 \pm 0.01	3.54 \pm 0.46	3.51 \pm 0.30	2.54 \pm 0.31	67 ^a
Cr	1.51 \pm 0.23	2.17 \pm 0.05	1.32 \pm 0.16	1.20 \pm 0.13	2.3 ^a
Pb	2.48 \pm 0.41	1.87 \pm 0.19	2.69 \pm 0.58	2.84 \pm 0.63	10 ^b
Cd	0.17 \pm 0.41	0.18 \pm 0.19	0.14 \pm 0.58	0.12 \pm 0.63	0.3 ^b

Key: ^aSource: [23], ^bSource: [24], NA = Not Available, sd = standard deviation.

3.3. Statistical Analysis

3.3.1. Analysis of Variance

Variations in the mean levels of metals between the samples were tested using one-way ANOVA. The results of metal concentration indicated that significant differences were obtained ($p < 0.05$) at 95% confidence levels for Na, K, Ca, Mg, Fe, Cu, Co, Ni, Cr and Pb in garlic bulb samples collected from all the four sites. However, the variations of Zn, Mn, Mo and Cd for the garlic bulb samples were not significant ($p > 0.05$). Similarly, significant differences were obtained ($p < 0.05$) in the levels of Na, K, Ca, Mg, Fe, Zn, Mn, Cu, Mo, Co, Ni, and Cd in garlic leaf samples collected from all the four sites. However, the variations of Cr and Pb for the garlic leaf samples were not significant ($p > 0.05$). This significance difference and increase in elemental concentrations may be due to application of various types of pesticides and fertilizer, the type of water present in the soil, and difference in physicochemical nature of the soil.

3.3.2. Pearson Correlation

The significant relationships between concentration of essential and non-essential metals in *Allium sativum* bulb and leaf samples were further substantiated by performing correlation analysis. Pearson's correlation coefficients of mean level of metals from all the sample sites ($n = 4$) between garlic bulb and garlic leaf was analyzed at $p = 0.05$.

According to literatures, if correlation coefficient is 1.0, there is complete dependency, if it is 0.0 there is no relationship, if it is negative, both are said to be correlated in opposite direction. However, if correlation is > 0.50 , it is said to be significant and less significant when < 0.50 [25].

The values of Pearson correlation coefficients between metal concentrations of garlic bulb and garlic leaf samples are given in Table 7. As can be seen from Table 7, statistically significant positive correlation coefficients for Na ($r = 0.758$), Ca ($r = 0.581$), Mg ($r = 0.965$), Fe ($r = 0.882$), Zn ($r = 0.862$), Mo ($r = 0.541$), Co ($r = 0.995$), Cr ($r = 0.889$) and Cd (0.587) were established between metal concentrations in garlic bulb and leaf samples.

From Table 7, there were weak positive correlations for K ($r = 0.192$) and Ni ($r = 0.448$). There were negative insignificant correlations for Mn, Cu and Pb between garlic bulb and leaf samples. The weak negative insignificant correlation indicates there was weak association between *Allium sativum* bulb and leaf samples at both locations.

Table 7. Correlation coefficient (r) for metals between garlic bulb and leaf.

Metals	r	Metals	r
Na	0.758	Cu	-0.151
K	0.192	Mo	0.541
Ca	0.581	Co	0.995
Mg	0.965	Ni	0.448
Fe	0.882	Cr	0.889
Zn	0.862	Pb	-0.917
Mn	-0.418	Cd	0.587

4. Conclusion

The levels of essential metals (Na, K, Ca, Mg, Fe, Zn, Mn, Cu, Mo, Co, Cr and Ni) and non-essential metals (Cd and Pb) in *Allium sativum* (garlic) bulb and leaf samples collected from Ambo Woreda, Ethiopia were determined for Na and K using flame photometer, Ca and Mg by EDTA titration, and the rest of the metals using ICP-OES after wet digestion. The optimized wet digestion method for digestion of the garlic samples were found to be efficient, precise and accurate for the metals analyzed, and it was validated through the recovery experiment and a good percentage recovery was obtained for the essential and non-essential metal determined.

This study revealed that the investigated garlic bulb and leaf samples are good source of essential metals. However, the results show that the levels of metal contents were higher in the leaves than the bulbs. It was generally observed that the results obtained are in agreement with the FAO/WHO guideline (2001) for allowed elemental concentrations in human nutrition: hence the garlic bulb and leaf samples can be consumed without any risk. Furthermore, the present study will give brief information about the mineral contents of garlic bulb and leaf, and these results may serve as a base line data for determination of mineral contents in vegetables in the study area.

In conclusion, it was found that various farming activities and heavy usage of fertilizers and pesticides did not increase the content of toxic metals in the study area. Awareness of people and regular monitoring of levels of these metals in vegetables is essential to prevent the incorporation in the food chain. Further works should be carried out in the soil samples where the vegetables are grown.

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References

- [1] Kamenetsky, R., Faigenboim, A., Mayer, E. S., Michael, T. B., Gershberg, C., Kimhi, S., Esquira, I., Shalom, S. R., Eshel, D., Rabinowitch, H. D. and Sherman, A. (2015). Integrated transcriptome catalogue and organ-specific profiling of gene expression in fertile garlic (*Allium sativum* L.). *BMC Genomics*, 16 (12), 1-15.
- [2] Kallel, F., Driss, D., Chaari, F., Belghith, L., Bouaziz, F., Ghorbel, R. and Chaabouni, S. E. (2014). Garlic (*Allium sativum* L.) husk waste as a potential source of phenolic compounds: Influence of extracting solvents on its antimicrobial and antioxidant properties, *Industrial Crops and Products*, 62, 34-41.

- [3] Diriba, S. G., Kebede, W., Nigussie, D. R., Getachew, T. and Sharma, J. J. (2013). Postharvest quality and shelf life of garlic bulb as influenced by storage season, soil type and different compound fertilizers. *Journal of Postharvest Technology*, 1 (1), 69-83.
- [4] Deresse, D. (2010). Antibacterial effect of garlic (*Allium sativum*) on *Staphylococcus aureus*: An *in vitro* study. *Asian Journal of Medical Sciences*, 2 (2), 62-65.
- [5] Packia Lekshmi, N. C. J., Viveka, S., Jeeva, S. and Raja Brindha, J. (2015). Antimicrobial Spectrum of *Allium* Species: A Review. *Indian journal of Science*, 15 (44), 1-5.
- [6] Adekunle, I. M., Olorundare, O. and Nwange, C. (2009). Assessments of lead levels and daily intakes from green leafy vegetables of southwest Nigeria. *Nutrition and Food Science*, 39 (4), 413-422.
- [7] Hellen, L. E., & Othman, O. C. (2014). Levels of selected heavy metals in soil, tomatoes and selected vegetables from Lushoto district-Tanzania. *International Journal of Environmental Monitoring and Analysis*, 2 (6), 313-319.
- [8] Kachenkom, A. G. and Singh, B. (2006). Heavy metal contamination in vegetables grown in urban and metal smelter contaminated sites in Australia. *Water, Air and Soil Pollution*, 169 (1-4), 101-123.
- [9] Wilson, B. and Pyatt, F. B. (2007). Heavy metal dispersion, persistence, and bioaccumulation around an ancient copper mine situated in Anglesey, UK. *Ecotoxicology and Environmental Safety*, 66, 224-231.
- [10] Tasrina, R. C., Rowshon, A., Mustafizur, A. M. R., Rafiqul, I. and Ali., M. P. (2015). Heavy metals contamination in vegetables and its growing soil. *Journal of Environmental Analytical Chemistry*, 2 (3), 1-6.
- [11] Chailapakul, O., Korsrisakul, S., Siangroh, W. and Grudpan, K. (2007). Fast and simultaneous detection of heavy metals using a simple reliable microchip-electrochemistry route: An alternative approach to food analysis. *Talanta*, 74, 683-689.
- [12] Hamza, N. A. E., Hammad, A.Y. and Eltayeb, M.A. (2013). Adsorption of Metals (Fe(II), Cr(III) and Co(II)) from aqueous solution by using Activated carbon prepared from Mesquite tree. *Science Journal of Analytical Chemistry*, 1(2), 12-20.
- [13] Dibofori-Orji, A. N. and Edori, O. S. (2015). Analysis of some heavy metals (Pb, Cd, Cr, Fe, Zn) in processed cassava flour (garri) sold along the road side of a busy highway. *Archives of Applied Science Research*, 7 (2), 15-19.
- [14] Kassa, B. and Hailay, K. (2014). Spectroscopic determination of trace metals (Mn, Cu and Ni) content in *Moringa oleifera*. *International Journal of chemical and Natural Sciences*, 2 (5), 141-144.
- [15] Chauhan, A., Mittu, B. and Chauhan, P. (2015). Analytical method development and validation: A concise review. *Journal of Analytical and Bioanalytical Techniques*, 6 (1), 1-5.
- [16] Iqbal, J., Carney, W. A., LaCaze, S. and Theegala, C. S. (2010). Metals determination in biodiesel (B100) by ICP-OES with microwave assisted acid digestion. *The Open Analytical Chemistry Journal*, 4, 18-26.
- [17] Kiflom, G. and Tarekegn, B. (2015). Determination of some selected heavy metals in fish and water samples from Hawassa and Ziway Lakes. *Science Journal of Analytical Chemistry*, 3(1), 10-16.
- [18] Shrivastava, A. and Gupta, V. B. (2011). Methods for the determination of limit of detection and limit of quantitation of the analytical methods: Review Article. *Chronicles of Young Scientists*, 2 (1), 21-25.
- [19] Thomas A. L. (2015). *Method validation essentials, limit of blank, limit of detection, and limit of quantitation*. Bio Pharma International, 28 (4), 48-51.
- [20] Mitra, S. (2003). *Sample preparation techniques in analytical chemistry* (Vol. 162, pp. 6-244). Hoboken: John Wiley and sons, Inc.
- [21] Harvey, D. (2000). *Modern analytical chemistry* (1st ed., pp. 706-7110). Depauw University, United States of America: McGraw-Hill.
- [22] USEPA. (2010). *National functional guidelines for inorganic superfund; Data review*.USEPA-540-R10-011, Washington, DC.
- [23] FAO/WHO (Codex Alimentarius Commission). (2001). Food additives and contaminants. *Joint FAO/WHO food standards program: ALINORM 01/12A: 1-289*.
- [24] WHO. (1998). *Quality control methods for medicinal plant materials*, WHO, Geneva, Switzerland.
- [25] Umar, M. A. and Salihu, Z. O. (2014). Heavy metals content of some spices available within FCT-Abuja, Nigeria. *International Journal of Agricultural and Food Science*, 4 (1), 66-74.