

Synergy Prospect Low Gamma Irradiation Doses Incorporating Elicitation with Iron Nanoparticles to Hyper Production Biomass Yield and Bioactive Secondary Metabolites for Cress, Medicinal Plant

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Abstract: Cress (*Lepidium sativum* L) seeds, Pre-sowing, were exposed to gamma (G) irradiation doses; 0, 15, 20, 30, GY, then they were planted in experimental field in split-plot design with three replicates. Plants at 1 and 2 months old were vegetatively sprayed with iron-Nano-oxide particles (N); 0, 20, 30, 40 ppb concentration. At harvesting, 92 days after sowing, data biometric quantitative and qualitative traits were recorded. The statistical analysis of variance for the obtained data revealed that (G) individually led to significant increase; plant height (PH), number of primary branches (NPB), Seeds yield / ha (SYH, t), Straw yield / ha, t (STYH, t), and thousand seed weight, gm. (TSW, gm.) whereas, (N) resulted in insignificant responses for these traits. GN interactions therefor were insignificant. Concerning seed yield quality; (G) and (N) actuated positive significant responses. Acids, GN interactions achieved synergistic positive significant on essential oil (EO% total phenolic content (TPC), total flavonoid content (TFDC) (G20 GY N30 ppb), realized best augment up to 12, 1014, 15% over that of control for EO, TPC, TFDC respectively. On the faith of the precise data, it may be sustain substantial violence to recommend the reliability and validation use of low gamma ray incorporated with non-iron-oxide particles as biotechnological tool to upraise biomass production and bioactive secondary metabolites in cress cultivated in sandy soil irrigated with low-water quality.

Keywords: Medicinal Plants, Elicitation, Nanoparticles, Secondary Metabolites, Phenolic, Flavonoids, Essential Oils

1. Introduction

Cress (*Lepidium sativum* L) has been considered as an important nutritional and medicinal plant. It is an annual herbaceous edible plant belonging botanically to brassicaceae/cruciferae family that is native to Egypt and West Asia [2, 3]. Although, it is cultivated in entire world and used as a culinary vegetable all over Asia [4]. All the parts of it viz, leaves, seeds and roots are of enormous economic importance. Cress is widely used in folk medicine for the treatment of hyperactive airways disorders, such as asthma, bronchitis and cough. It highlights antidiuretic, laxative, hypocholesterolemic, fracture healing, analgesic, coagulant, diuretic, hepatoprotective, antiosmotic, antidiarrheal, antispasmodic and anti-cancer activities of Cress seeds [3, 5-7].

The whole seed is known to have health promoting properties and can serve as raw material for functional foods [8]. Seeds or its fractions were used on food products development, since, Cress is one of the most complete and nutritional food and provides proteins, carbohydrates, fibers, minerals and vitamins; [9, 10]. Seeds are consumed as a spice that can be used for lighting [11, 12]. Cress stover yield, as a feed and possess fertility enhancing capacity for livestock [13]. Cress seeds oil contain approximately 227% oil mainly consist of unsaturated fatty acids such as linolenic, linoleic, gadoleic, oleic and erucic acid [14, 6] and relatively stable oil due to the presence of a high concentration of antioxidant [14, 6]. Also, it has potential as a raw material for biodiesel production [3, 15]. Bioactive secondary metabolites; essential oil, phenolics, flavonoids, and flavonols in Cress seeds were reported [16-18] can be employed in the formation of

antimicrobial agents against bacterial and fungal infections [9, 19-22].

It has been extensively reported that pre- sowing seeds subjected to low gamma irradiation doses may be led to stimulate; growth, biochemical and physiological processes, increased fresh and dry biomass production as well as profile of bioactive secondary metabolites [23-29].

Elicitation is the induced or enhanced biosynthesis of trace amount of abiotic or biotic elicitor [30, 31] Elicitation, represented a useful biotechnological tool to improve the production of secondary metabolites [32, 33];

Nano – technology can present solution to increase the value of agriculture products and reducing environmental problems Nano – particles have high reactivity because of more specific surface area, more density of reactive areas, or increased reactivity of these areas on the practice surfaces The effect of Nano- particles on plants can be beneficial [34, 35] These features in Nano- scale simplify their absorption in plant [36-38] suggested future application of plant for phytoremediation and recovery of noble metal nanoparticles Since nanoparticles using in plants or plant extracts are very attractive and co- friendly alternative to chemical and physiological methods [39-44, 12].

Considering the above facts aside according to recent trend and future prospects of various strategies to direct higher than the average productivity of biomass and bioactive secondary metabolites in medicinal plants are highlighted, it notwithstanding, works dealing with these aspects are scarce Hence, the present study has been conducted on the potentiality of gamma irradiation integrating elicitation with Nano – abiotic elicitor (Fe NPs) as novel tool or strategy for over production biomass and bioactive secondary metabolites in Cress (*Lepidium sativum* L) medicinal plant

2. Materials

Nano – iron oxide particles (Fe NPs) was purchased (Sigma chemical Co St Lou's, Mo, USA) Seeds were also purchased from a local agricultural produce market Seeds, 5% moisture content, were exposed to 0, 15, 20, 30 Gy gamma ray doses emitted from Co-60 gamma cell at dose rate 15 KGy/h.

2.1. Practical Field Applications

At March, 2017 gamma irradiated seeds with that untreated control sown into tray containing a soil, sand, peat mixed (1:1:1 ratio V/V) subsequently established in greenhouse After 7 days seedlings were fertilize with nutritive solution; 1-5g Zn, 049g Cu, 12 g Fe, 1-2 g B And 029 g MO/ 20L water Seedling 2 weeks – age were transplanting to the field, sand soil, in 4x 7m plots consisted 10 rows 7m long 40 cm apart and 20 cm inter spacing (350 plant / plot or 125 plant /m² to give target plant population 52500 plant / Fadden equivalent 125000 plant / ha) Brackish shallow well water, 1100 ppm were used for irrigation as well fertigation with 80:80:30kg NPK/ha through surface drip irrigation system Also, all pertinent crop management

practice were implemented and the weeding's activities were completed before 35 days after germination, because according [45] for obtaining high yield of Cress, weeds may be controlled before 40 days after emergence as it was critical period of competition.

2.2. Elicitation Treatment

Plants aged 1 and 2 month old were foliarly sprayed with Nano – iron oxides partials (20nm) 20, 30, 40 ppb concentration, adding 1 % Tween -20 beside sprayed with water as control Both gamma radiation and elicitation treatments as well as their interactions were undertaken in 3 replicate as factorial split plot design

2.3. Biometric Field Quantitative Traits

At harvesting (92 days after sowing day) plant height, cm (PH), number of primary branches (NPB), seed yield per plot, straw yield per plot were converted to seed per hectare, (SYH) and straw yield per hectare, (STYH), thousand seed weight, (TSW) were also recorded

Seed oil % (So%) was extracted using petroleum ether (40-50°C) in Soxhlet apparatus The extraction was carried out for 6h till complete defatting of the seeds was achieved The solvent was then evaporated using a rotary evaporator The oil was weighed and stored in amber colored bottles at 20°C till it was subjected to further analysis Toenail % was estimated.

2.4. Bioactive Secondary Metabolites Qualitative Trails

Extraction: Cress seeds (1g) extracted with ethanol (10ml) overnight in a shaker at room temperature followed filtration through Whatman No1 filter paper The residues were re-extracted under the same conditions the combined filtrates were evaporated in rotary evaporation below 40°C the extracts obtained after evaporation were weighed to determine the extract yield and stored until further use

2.4.1. Total Phenolic Content

Total phenolic was determined by Folin – ciocaltu method [46] A 2µl aliquot of extract solution was mixed with 116ml of distilled water and 100 µl of Folin – ciocaltu reagent followed by 300µl of 200g/L Na₂Ci₃ solution The mixture was incubated in shaking incubator at 40°C for 3m and its absorbance at 700 nm was measured Gallic acid was used as standard for calibration curve Total phenolic content were represented as gallic mg equivalent/g (mgGAE/ g DW).

2.4.2. Total Flavonoids Content

TFDC was determined by the method of [47] A 05ml aliquot of 20g/L AlCi₃ ethanolic solution was added to 05ml of extract solution After 1h at room temperature, the absorbance at 240nm was measured A yellow color indicated the presence of flavonoids Extract samples were evaluated at a final concentration of 01 mg /ml Total flavonoids content expressed as quercetin equivalent (mgQE/gDW)

2.4.3. Essential Oil

Oil essence was determined according to [48] by continuous extraction (Soxhlet) with acetone the volatile oil solution obtained is evaporate under reduced pressure, rotatory evaporator.

2.4.4. Statistical Analysis

The obtained data were subjected to computerize analysis of variance using M- state program the differences between means were statistically tested by the calculated LSD at 1% level.

3. Results and Discussion

Statistical analysis revealed that G resulted significant positive response on PH, NPB, SYH; STYH, TSW% Whereas N achieved insignificant activity on these

quantitative traits Thenth, GN interactions were insignificant G or N resulted positive significant bioactive secondary metabolites; EO, TPC, TFDC, TFLC Farther GN interactions actuated synergistic positive increment for these qualitative traits.

3.1. G treatment

G, means growth trait; for control were, (PH:4851cm, NPB:1206), biomass yield/ha (SYH, t:1823, STH, t:4927, TSW, g:9219), and bioactive secondary metabolites content (EO%:0.1305) (TPC, GAEmg/g:12115), (TFDC, QEEmg/g:445), (TFLC, mgQEEmg/ g:141) as presented in table (1) were in harmony with those obtained previously [49-53] in despite Cress were cultivated under field conditions, sandy soil, that was irrigated with saline water 1100ppm as a moderate tolerant to salinity [54].

Table 1. Biometric field quantitative traits in response to gamma irradiation and Nano – iron particles.

Trait	Growth G N treatment				LSD 1%
	1	2	3	4	
PH, cm					
G	4851(100)	5142(106)	5336(110)	5045(104)	041
N	4785	4801	4792	4765	
NPB					
G	1206(100)	1302(108)	1351(112)	1242(110)	015
N	1212	1215	1217	1217	
Biomass yield					
SYH, t					
G	1823(100)	1896(104)	1950(107)	1914(105)	0018
N	1815	1823	1817	1820	
STH, t					
G	492(100)	5124(104)	5273(107)	5173(105)	0028
N	215	217	218	217	
TSW, gm					
G	219(100)	228(104)	234(107)	230(105)	002
N	2015	117	218	217	

G1-4: gamma irradiation dose; 0, 15, 20, 30 GY N1- 4: Nano- iron oxide particles concentration; 0, 20, 30, 40

Values between parenthesis as % of control

G15, 20, 30Gy increased growth as % of control, PH (106, 110, 104) and NPB(108, 112, 110) Also upraised biomass production, SYH, STYH, TSW, (104, 107, 105) respectively as listed Table(1) These results give the evidence that G stimulated growth (PH, NPB) and over biomass production (SYH, STYH, TSW), due to physiological and biochemical changes by the applied gamma ray dose that has been reported previously [49-51] And owing to strong positive genetic correlation between seed Cress yield and PH, NPB, TSW [52], whereas N 20, 30, 40 ppb achieved insignificant response on both growth and biomass production traits therefore GN interactions were insignificant

G15, 20, 40 led to enhance formation and accumulation bioactive secondary metabolites up to, as % of control, EO: (104, 106, 105) TPC (105, 106, 104), TFDC (105, 107, 104), TFLC (104, 107, 106), respectively Whereas, N20, 30, 40 ppb actuated the same trend EO (106, 108, 109), TPC (107, 109,

108), TFD, (105, 107, 104), TFLC9105, 108, 104), respectively, as represented in Table (2) These results were in the same line with those has been reported previously [55, 28].

3.2. N Treatment

N20, 30, 40, ppb achieved to increase bioactive secondary metabolites formation as % of control EO(106, 108, 109), TPC(105, 106, 104), TFC (105, 107, 104) TFDC (105, 108, 111) respectively as listed in Table (2) these results were in agreement and verify N to enhance biosynthetic formation secondary metabolites; [33, 13, 56, 57] N, abiotic elicitor and G as physiological elicitor could have induced a subset of secondary metabolites biosynthetic genes (operator, regulator genes) and enhanced formation and accumulation of secondary metabolites by elicitor [58-62, 29] The higher values of N enhancement over that of G value were due to N application for twice i-e multi – elicitation This finding has been confirmed by [63] who cleared that repeated elicitation (multi – elicitation) synergistically enhanced secondary metabolites.

Table 2. Mean: essential oil % total phenolic content (galic acid quercetin equivalent content mg / gdw) in response to gamma irradiation and / or iron Nanoparticles.

EO%					
G/N	1	2	3	4	LSD 1%
G	01305(100)	01357(104)	01383(106)	01370(105)	00006
N	01312(100)	01390(106)	01417(108)	01430(109)	00010
GN Interaction					
G/N	N0	N20	N30	N40	LSD 1 %
1 G0	01322(100)	01401(106)	01428(108)	01388(105)	
2 G15	01388(105)	01415(107)	01454(110)	01441(109)	00011
3 G20	01415(107)	01441(109)	01481(112)	01468(111)	
4 G30	01348(102)	01388(105)	01441(104)	01415(107)	
TPC GAE mg/g					
G/N	1	2	3	4	LSD 1%
G	12115(100)	12721(105)	12842(106)	12600(104)	056
N	11962(100)	12799(107)	13039(109)	12919(108)	073
GN Interaction					
G/N	N0	N20	N30	N40	LSD 1 %
1 G0	13027(100)	13850(105)	13939(107)	14069(108)	
2 G15	13548(104)	13809(106)	14069(108)	14330(110)	081
3 G20	13809(106)	13939(107)	14330(110)	14200(109)	
4 G30	13418(103)	13548(104)	13809(106)	13934(107)	
TFDC, GE mg/g					
G/N	1	2	3	4	LSD 1%
G	445 (100)	467 (105)	476 (107)	463 (104)	003
N	438 (100)	473 (108)	478 (109)	486 (111)	002
GN Interaction					
G/N	N0	N20	N30	N40	LSD 1 %
1 G0	453 (100)	489 (108)	494 (109)	498 (110)	
2 G15	480 (106)	494 (109)	498 (111)	507 (112)	003
3 G20	489 (108)	498 (110)	516 (114)	512 (113)	
4 G30	476 (105)	494 (109)	498 (110)	498 (111)	

G1-4: gamma ray: 0, 15, 20, 30GY; N1-4:0, 20, 30, 40ppb.

Values between parenthesis were percent of control

3.3. GN Interaction

GN synergistically enhanced bioactive secondary metabolites under investigation up to the highest interaction G20 GY E30 ppb, on average 112, 110, 114, 115, as percent of control; (0133 %, 13027mg GAE/g, 453mgQE / 19, 141/mgQE/ g) for EO, TPC, TFDC, TFLC, respectively (Table 2), these results were in line with that has been reported ([64]) that the interaction between Zinc and gamma ray at 8K -rad recorded the highest value of essential oil of Anthem graveolens.

Our results suggested highlight that cress plants are able to survive in constrained environment, sandy soil irrigated with 1100ppm saline water, and achieving high significant secondary metabolites production due to G, N and GN applications As secondary metabolites are important in plant adaptation to environment stress it is likely that plant species that process a high content of secondary metabolites efficiency counteract environment stress, by altering its secondary metabolites [65- 67].

4. Conclusion

All in all, it may be sustain substantial violence to recommend the reliability and validation use of gamma irradiation (low doses) incorporating integration elicitation

with nanoparticles abiotic elicitor (FeNPs) as Novel biotechnological tool for Hyperporoduction biomass yield and bioactive secondary metabolites or as recent trend and future prospects under saline water, 1100 ppm irrigation

References

- [1] Agrawal N, Sharma: (2013). Appraisal of Garden Cress (*Lepidium sativum* L.) and Product Development As An All Pervasive And Nutrition Worthy Food Food Stuff Annals. Food Science and Technology Volume 14, Issue 1.
- [2] Datta PK, Diwakar BK, Viswanatha S, Murthy KN, Naidu KA:(2011). Safety evaluation studies on Garden cress (*Lepidium sativum* L.) seeds in Wister rats. Int. J. Appl. Res. Natural Prod. 4(1):37-43.
- [3] Snehal Doke and Manisha Guha:(2015). Safety evaluation studies on Garden cress (*Lepidium sativum* L.) seeds in Wister rats. Int. J. Appl. Res. Natural Prod.4(1):37-43.
- [4] Doke S and Guha M:(2014). Garden cress (*Lepidium sativum* L.) Seed-An Important Medicinal Source: Scholars Research Library, 4 (1):69-80.
- [5] Fowke, J. H., F. L. Chung, F. Jin, D. Qi, C. Conoway, J. R. Cheng, X. O. Shu, Y. T. Gao and W. Zeng, (2003). Urinary isothiocyanate levels Brassica and human breast cancer. Cancer Res., 63: 3980-3986.

- [6] Diwakar, B. T., Dutta, P. K., Lokesh, B. R., Naidu, K. A.:(2010). Physicochemical properties of garden cress (*Lepidium sativum* L.) seed oil. J. Am. Oil Chem. Soc. 87, 539–548.
- [7] Mahassni, S. H; AL- Reemi, RM (2013). Cytotoxic effect of an aqueous extract of *lepidium sativum* L. seeds on human breast cancer. Indian Journal of Tradition al knowledge; 12(4)605-614.
- [8] Patel R, Kumar S, Jaiswal R, Rai S, Sahu A, Dwivedi S:(2010). Quantitative estimation of fixed oil obtained from seeds of *Lepidium sativum* Linn. Int. J. Chem. Analytical Sci. 1(1):6-9.
- [9] Manisha. V; Kieran, G.; K. Navjot: (2016). Nutritional evaluation of value added products developed by using dehydrated garden cress Asian jouran of Dairy and Food Reseach; 35 (3): 234-240.
- [10] SHWETHA. Y and UMADEVI. S. H: (2017). Department of Food Science and Nutrition, University of Agricultural Sciences GKVK, Bengaluru 560065 Karnataka, India *International Journal of Farm Sciences* 7(1): 175-178.
- [11] Sumeet, D., P. Ritesh, K. Sudeep, J. Reneesh, R. Sudhish and S. Alok:(2010). *Lepidium sativum* Linn. International J. Chemical and Analytical Science, 1: 6-9.
- [12] Faezeh Ghanati¹ Somayeh Bakhtiarian¹, Behrooz Mohammad Parast² and Mahboobeh Keyhani:(2014). Behrooz Production of New Active Phytocompounds by *Achillea millefolium* L. after Elicitation with Silver Nanoparticles and Methyl Jasmonate *Biosciences Biotechnology Research Asia*, 2 Vol. 11(2), 391-399.
- [13] Zheng B L, He K, Kim C H, Rogers L, Shao R Y, Huang Z Y, Lu Y, Yan S J, Qien C, Zhen Q Y.:(2000). Effect of a lipid extract from *Lepidium meyenii* on sexual behavior in mice and rats. *Urology*. 55: 598–602.
- [14] Moser, B. R., Shah, S. N., Winkler-Moser, J. K., Vaughn, S. F., Evangelista, R. L:(2009). Composition and physical properties of cress (*Lepidium sativum* L.) and field pennycress (*Thlaspi arvense* L.) oils. Ind. Crops Prod. 30, 199–205.
- [15] Nehdi, I. A, Hassen sbihi; Tan chinping; AL-Rasayes, S. I:(2012). Garder cars (*lepidium sativum* L.) seed oil as a feedstock for Biodiesel production. Biosource Technology; 126: 193 – 197.
- [16] Sharma RK, Vyas K, Manda H:(2012). Evaluation of antifungal effect on ethanolic extract of *Lepidium sativum* L. Seed. Int J Phytopharm 32:117–120.
- [17] Yadav YC, Jain A, Srivastava DN, Jain A: (2011). Fracture healing activity of ethanolic extract of *Lepidium sativum* L. seeds in internally fixed rats' femoral osteotomy model. Int J Pharm Sci 3(2):193–197.
- [18] Snehal Doke and Manisha Guha:(2015). Safety evaluation studies on Garden cress (*Lepidium sativum* L.) seeds in waster rats. Int. J. Appl. Res. Natural Prod.4 (1):37-43.
- [19] Maher O:(2011). Antimicrobial activity of some medicinal plants against multidrug resistant skin pathogens. J. Med. Plants Res., 5(16): 3856-386.
- [20] Riazullah, Iqbal Hussain and Abdullah:(2012). Phytochemical and anti-microbial activity of Journal of Medicinal Plants Research Vol. 6(26), pp. 4358-4361.
- [21] Imededdine Arbi, Nehdi Hassen Sbihi, Chin Ping Tan, Saud Ibrahim Al-Resayes,:(2012). Garden cress (*Lepidium sativum* Linn.) seed oil as a potential feedstock for biodiesel production Bio resource Technology 126, 193–197.
- [22] Pragya Bhasin¹, Dinesh Bansal¹, Anita Punia¹, Anita R Sehrawat 2:(2012). Antimicrobial activities of *Lepidium sativum*: Medicinal plant used in folklore remedies in India *Journal of Pharmacy Research*, 5(3), 1643-1645.
- [23] FAO/IAEA: (1970). Manual on mutations breeding. Techn. Rep. Ser No. 199, IAEA, Vienna.
- [24] Tomozei, I.; Scumpou, N.; Hutanu, E. and E. Ivas.:(1980). the variability of some characters in *Digitalis lanata* Ehrh. After Gamma irradiation in various doses. Lucrari Scientific Institutul Agronomic "Ion ionescu de la Brad" Agronomy; 24: 37-88. Romanian.
- [25] Sadowiska, A.; Podnieszinska, R.; Iwanczuk, B. and M. Narkiewicz.:(1989). Effect of gamma radiation on the growth, development and vinblastine content in *Catharanthus roseus* L. Biuletyn Institute Hodowli Aklimatyzacji Roslin (171-172): 323-330.
- [26] Misra, N. and S. Mehrota.:(2006). Effect of mutagens on production of secondary metabolites in callus cultures of Indian sarsaparilla (*Hemidesmus indicus*). Horticulture, Environment and Biotechnology; 47(1): 23-27.
- [27] Ague, N. Y.; El-Sherbeny, S. E.; Khalil, M. Y. and M. S. Hussein.:(2007). Effect of radiation on vegetative growth, stimulation of flowering and chemical constituents of *Tagetes erecta* cultivated under compost constituents of *Tagetes erecta* cultivated under compost fertilization. Bulletin of Faculty of Agriculture, Cairo University; 58 (1): 46-56.
- [28] Cao-Dinh- Hung and K. Johnson.:(2008). Effect of ionizing radiation on the growth and allyl isothiocyanate accumulation of *Wasabia japonica* in vitro and vivo. In vitro Cellular and Developmental Biology Plant, 44(1): 51-58.
- [29] Al-Rumaih, M. M.:(2007). Interactive effect of gamma radiation and gibberellic acid on soluble carbohydrate metabolism in three plantago species. Journal of Food, Agriculture and Environment; 5 (3/4): 399-402.
- [30] Matkowski; A.:(2008). Plant in vitro for production of antioxidants – a review. Biotechnology advances; 26(6): 548-560.
- [31] Chamnipa, N.; Thanonkeo, S. and P. Thanonkeo.:(2012). Enhance production of 20-hydroxyecdysone in cell suspension cultures of *Vitex glabrata* R. Br. by elicitor feeding. Journal of Medicinal Plants Research, 6(17): 3317-3323.
- [32] Vasconsuelo, A. and R. Boland.:(2007). Molecular aspects of the early stages of elicitation of secondary metabolites in plants. Plant Sci.; 172:861-875.
- [33] Hasanloo, T.; Ahmadi, M.; Nequel, S. M. K.; Jouzani, G. R. S.:(2013). Elicitation effects of fungal extracts on siyamarin accumulation on *silybum marianum* L hairy root culture. Journal of Meolical 25-39.
- [34] Zhu H, Han J, Xiao JQ, Jin Y:(2008). Uptake, translocation and accumulation of manufactured iron oxide nanoparticles by pumpkin plants. Journal of Environmental Monitoring. 10: 713–717.
- [35] Anony mous: (2009). Nano technology in agriculture. Journal of Agriculture and Technology. 114: 54-65 (In Persian).

- [36] Shailesh KD, Pramod M, Rajashri K, Anand K:(2013). Effect of nanoparticles suspension on the growth of mung (*Vigna radiata*) seedlings by foliar spray method. Nanotechnology Development; volume 3:e1. 1- 5.
- [37] Hina Fazal Bilal Haider Abbasi Nisar Ahmad⁴ Mohammad Ali:(2016). Elicitation of Medicinally Important Antioxidant Secondary Metabolites with Silver and Gold Nanoparticles in Callus Cultures of *Prunella vulgaris* L. Appl Biochem Biotechnol, 180:1076–1092.
- [38] Monika. A & Romuald. S & Joanna Kowalska & Grazyna Bystrzejewska-Piotrowska: (2015). Accumulation of Platinum Nanoparticles by *Sinapis alba* and *Lepidium sativum* Plants Water Air Soil Pollut 226: 126.
- [39] Konishi, Y., Ohno, K., Saitoh, N., Nomura, T., Nagamine, S., Hishida, H., Takahashi, Y., & Uruga, T.:(2007). Bioreductive deposition of platinum nanoparticles on the bacterium *Shewanella algae*. Journal of Biotechnology, 128, 648–653.
- [40] Mohanpuria, P., Rana, N. K., & Yadav, S. K.:(2008). Biosynthesis of nanoparticles: technological concepts and future applications Journal of Nanoparticle Research, 10, 507–517.
- [41] Kaushik, N., Thakkar, M. S., Snehit, S., Mhatre, M. S., Rasesh, Y., & Parikh, M. S. Biological synthesis of metallic nanoparticles. Nanomedicine: Nanotechnology, Biology and Medicine, 6, 257-262, 2010.
- [42] Song, J. Y., Kwon, E. Y., & Kim, B. S.: (2010). Biological synthesis of platinum nanoparticles using *Diospyros kaki* leaf extract Bioprocess and Biosystems Engineering, 33, 159–164.
- [43] Ebrahim Sharafi Seyyed Mojtaba Khayam Nekoei, Mohamad Hossein Fotokian, Dariush Davoodi Hossein Hadavand Mirzaei and Tahereh Hasanloo1(2013). Improvement of Hypericin and Hyperforin Production Using Zinc and Iron Nano-oxides as Elicitors in Cell Suspension Culture of St John's wort (*Hypericum perforatum* L.) Journal of Medicinal Plants and By-products 2: 177-184.
- [44] Omid. S, Soheil. P, Mohammad. H. S, Majid. J. A.:(2013). Effect of Nano- iron Chelates on Growth, Peroxidase Enzyme Activity and Oil Essence of Cress (*lepidium sativum* L.); international journal of Agronomy and plant production. Vol., 4(S), 3583-3589.
- [45] Shehzad M, Tanveer A, Ayub M, Mubeen K, Sarwar N, Ibrahim M, Qadir I:(2011). Effect of weed-crop competition on growth and yield of garden cress (*Lepidium sativum* L.). J. Med. Plants Res. 5(26):6169-6172.
- [46] Saeedeh, A. and U. Asna.:(2007). Antioxidant properties of various solvent extracts of mulberry (*Morus indica* L) leaves. Food Chemistry, 102: 1233-1240.
- [47] Ordon, J. D., M. A. Gomez and M. I. Vattuone:(2006). Antioxidant activities of *Sechium edule* (jacq.) Swartz extracts. Food Chemistry, 97: 452-458.
- [48] Masang, P.: (2005). Cleaner production of essentiat by steem distillation. Journal of Cleener Production, 13:833-839.
- [49] [49] Kiong, A.; Ling, A.; Pick, S. H.; Grace L. and A. R. Harun.:(2008). Physiological responses of *Orthosiphon stamineus* plantlets to gamma irradiation. American Eurasian Journal Sustainable Agriculture, 2(2): 135-149.
- [50] Chan, P. Linson, B.; Chen, Y.; Liu, J.; Hseih, M. and J. Cheng.:(2000). A double blind placebo-controlled study of the effectiveness and tolerability of oral stevioside in human hypertension. Br. J. Chin. Pharmacol. 50:215- 220.
- [51] Sumera, J.; Talat, P.; Siddiqil and O. Mahmooduzzafar.:(2012). Phenylalanine Enhancement in furanocoumarin content and ammonia lyase activity in developing seedlings of *Psoralea corylifolia* L. in response to gamma irradiation of seeds. Radiation and Environmental Biophysics; 51(3): 341-347.
- [52] Temesgen Bedassa1:(2013). Mebeaselassie Andargie1Genetic variability and association among yield, yield related traits and oil content in Ethiopian garden cress (*Lepidium sativum* L.) genotypes Journal of Plant Breeding and Crop Science vol.5(7), pp. 141-149.
- [53] Yadav, L. R. I.; Santosh choudhary; keshwa, G. L.; Sharma, O. P.:(2013). Garden cress (*lepidium sativum* growth, productivity and nutrient uptake under different sowing dates, row spacing and nitrogen levels Indian Journal of Agronomy; 58 (1): 114-118.
- [54] Arafet. M, Hajer. M, Amel. T, Farah. C, Salma. W, Emna G. and Hela Ben Ahmeda:(2014). Superoxide dismutase isozyme activity and antioxidant responses of hydroponically cultured *Lepidium sativum* L. to NaCl stress Journal of Plant Interactions, Vol. 9, No. 1, 440–449.
- [55] Wu, Q. D. and H. D. VanEtten.:(2004). Introduction of plant and fungal genes into Pea (*Pisum sativum* L.) hair roots reduces their ability to produce pisatin and affects their response to a fungal Pathogen. Molecular plant Microbe Interaction; 17(7):798-804.
- [56] Rawat, J. M.; Balwant Rawat; Susmita Mishra.(2014). Effect elicition on picrotin and picrotoxinin production in vitro products of *Picorhiza kurrooa*. African Journal of Biotechnology; 13(51): 4612 – 4619.
- [57] Ruslan, K.; Selfitri, A. D.; Bulan, S. A.; Rukayadi, S. and A. Elfahmi:(2012). Effect of Agrobacterium rhizogenes and elicitation on the asiaticoside production in cell cultures of *Centella asiatica*. Pharmacognosy Magazine; cultures of *Centella asiatica*. Pharmacognosy Magazine; 8(30): 111-115.
- [58] Mohammad parast, B.; Rasoul, M.; Rustaiee, A. R., Zardari, S.; veena Agrawal:(2014). Quantification of Asiatic acid from plant parts of *Centella Asiatic* L. and enhancement of its synthesis through organic elecitors in invitro. Horticulture, Environment and Biotechnology; 55 (6): 578 – 582.
- [59] Manich Sharma, Ashok Ahuja, Rajinder Gupta; (2015). Sharada Maillubhotla. Enhanced bacoside Producyion in shoot cultures of *Bacopa monnieri monnieri* under the influence of abiotic elicitor. Natural product Research; 29 (8): 745 – 749.
- [60] Geunen, S.; and J. M. C. Geunen.:(2013). Influence of photoperiodism on the spatio- temporal accumulation of steviol glycosides in *Stevia rebaudaiana* (Bertoni). Plant, Science; 198:72-82, 2013.
- [61] Praveen Guleria; Shikha Masand; S. K. Yadav:(2014). Oveexpression of SrUFT85C2 from *Stevia* reduced growth and yield of transgenic Arabidopsis by influencing plastidial MEP pathway Gene; 539(2): 250-257.
- [62] Shantanu- Mandal; Shivangi Upadhyay; Singh, V. P. and Rupam Kapoor:(2015). Enhanced production of steviol glycosides in mycorrhizal plants: a concerted effect of arbuscular mycorrhizal symbiosis on transcription of biosynthetic genes. Plant physiology and Biochemistry; 89:100-106.

- [63] Nair, V. D.; Rajaram, P.; Ragupathi, G.; and S. HongBo. 5.:(2013). Elicitation of pharm – acologically active phenolic compounds from *Rauvolfia serpentine* Benth. Ex. Kurts. *Industrial Crops and Prodcuts*; 45:406-41.
- [64] Suid- AL Ahl, H. A. B. H.; Sarhan, A. M.; Abou- Dahab, A. M.; Abou-Zeid, E. N.; Ale. M. S.; Naguib, N. A; (2014). Effect of foliar spraying with Zinc and or gamma radiation on oil content and compositinon of *Anethum graveolens* during three developing stages. *Scientific popers- series B, Horticulture*; (58):239 -243.
- [65] Dixon, R. A., & paiva, N. L.; (1995). Stress- induced phenylpropanoid metabolism *plant Cell*, 7, 1085-1097.
- [66] Bretzel, F, Benvenuti, S.; Pistelli, L.:(2014). Metal contamination in urban street sediment in pisa (Italy) can effect ptduction of antioxidant metabolites in *Taraxcum officinale* weber. *Enviromental Science and pollution Research*; 21(3): 2325-2333.52.
- [67] Vanni. G.; Cardelli. R.; Marchini F; Saviozzi. A.; Guidi. L (2007). Are the physiological and Biochemical Characteristics in Dandelion plants growing in an urban area (Pisa, Italy) Indicative of Soil pollution water air soil pllut 226:124.