

---

# Histopathological Study of the Rat Liver Exposed with Lead Acetate as a Microscopic Survey

Khatere Khosravian Dehkordi<sup>1, \*</sup>, Soraya khosravian Dehkordi<sup>2</sup>, Rahmat Allah Fatahian Dehkordi<sup>3</sup>

<sup>1</sup>Department of Animal Physiology, Islamic Azad University, Shahrekord Branch, Shahrekord, Iran

<sup>2</sup>Department of Basic Sciences, Azad University of Shahrekord, Shahrekord, Iran

<sup>3</sup>Department of Basic Sciences, Faculty of Veterinary Medicine, University of Shahrekord, Shahrekord, Iran

## Email address:

dehkordi\_khosravian@yahoo.com (K. K. Dehkordi)

## To cite this article:

Khatere Khosravian Dehkordi, Soraya khosravian Dehkordi, Rahmat Allah Fatahian Dehkordi. Histopathological Study of the Rat Liver Exposed with Lead Acetate as a Microscopic Survey. *Animal and Veterinary Sciences*. Vol. 3, No. 5, 2015, pp. 141-143.

doi: 10.11648/j.avs.20150305.14

---

**Abstract:** Lead is one of the environmental pollutants that can effect on the life of living animates in several ways; it has a long half-life and is collected in the soft tissue and conduct to adverse effects in tissues. The present study was performed to investigate the histological effects caused by lead in the rat liver. The study was conducted on 20 rat, the animals were divided into 2 equal groups. The first group received distilled water and considered as a control group. The second groups were orally administered lead acetate 8.5 mg/l of body weight for 20 weeks. The rats were anesthetized, the liver were removed for histological studies. Histological changes which observed in the liver were vacuolation, fatty degeneration, congestion within central veins, hemorrhage and infiltration of inflammatory cells. In this study, harmful toxic effects observed in liver of rats.

**Keywords:** Lead, Histological Alterations, Liver

---

## 1. Introduction

For years, lead had been a toxic problem for human and animals [1] and the most useful and complete model for toxicological studies is the toxicity of lead [2-4]. In spite of its detected hazards, this element used widely at industrial and commercial applications as in paints, plumbing, manufacture of lead acid batteries and etc. [5, 6]. Gastrointestinal and respiratory systems are more exposed to lead and therefore create acute and chronic situations [7]. When foods are put into improperly glazed pottery or ceramic dishes, lead can enter foods. Lead also can leach into drinking water in water distribution systems of individual house [8, 9]. Soft tissues and bone are place that the ingested and absorbed lead was stored and the highest lead concentration occurs within the teeth, bone, lung, liver brain, kidney and spleen [10].

The histomorphometrical properties of the hepatic tissue following lead intoxication are not evidenced. This study was performed to characterize the possible histomorphometric changes in the hepatic tissues following experimental lead poisoning in rats.

## 2. Materials and Methods

### 2.1. Animals

Twenty adult male rats (*Rattus norvegicus albinus*) that were 1.5 months old, weighing 210±35 g, were used in this study. The animals were maintained in individual cages at the room with a 12-h light and 12 dark cycles and limited temperature of 20±1°C, and were permitted access to water and standard laboratory pellets ad libitum. The assay protocol for this research was approved by the University Research Committee.

### 2.2. Experiment Protocol

Rats were randomly divided into two groups, A and B; n=10 each group. The animals in group A served as the control group and received the distinct dosage of distilled water. The group B received 8.5 mg/l by oral gavage of the lead acetate (fulda-Germany). Experiment design was followed for 20 consecutive days in all groups.

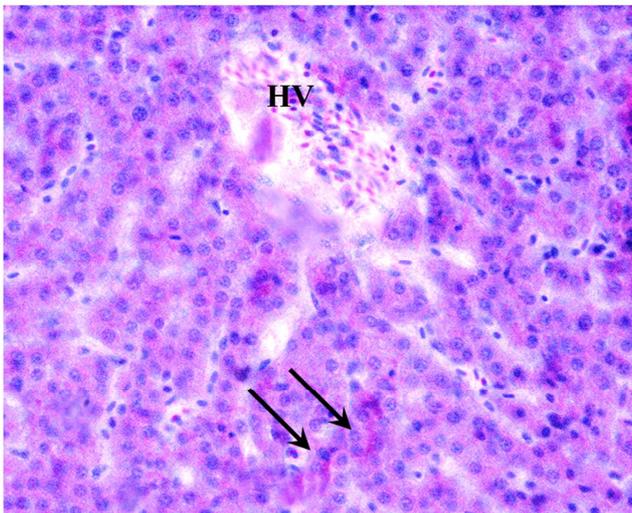
### 2.3. Tissue Processing

Portions of the lobes of the liver from each rat were cut

rapidly, fixed in neutral buffered formalin (10%), were then dehydrated, with grades of ethanol (70%, 80%, 90%, 95% and 100%). Cleared in xylene and embedded in paraffin wax, then 4-5  $\mu\text{m}$  thick sections were obtained by rotary microtome and stained with Harris hematoxylin and Eosin (Luna, 1968).

### 3. Results

The lobular engineering into the liver of lead-administered rats was kept intact in all treated rats after 20 consecutive days of 8.5 mg/l the lead acetate administration. In comparison with the control group, in some hepatocytes was seen vacuolation, fatty degeneration in other hepatocytes, congestion within central veins and some sinusoids. Infiltration of inflammatory cells was observed in the interstitial tissue in addition to the congestion of the central veins (fig 1).



**Figure 1.** Histopathological photograph of rat liver in experimental group; congestion within central veins (Hv) and some sinusoids (arrows); (H&E,  $\times 200$  magnification).

The sinusoidal Kupffer cells showed a prominent appearance and therefore were increased in number following lead intoxication.

In control group was showed normal lobular architecture and histology. In control group; the central vein, the initial branch of hepatic vein, was occupying the longitudinal axis of each classical lobule. Hepatocytes radiated from the central vein fenestrated plates of liver cells, separated from each other by large vascular spaces known as hepatic sinusoids.

### 4. Discussion

Lead element is one of the most common toxic metals and the most common route of exposure is by ingestion of lead contaminated food [11]. Once it is absorbed from gastrointestinal tract, lead bounds to erythrocytes and widely distributed initially to soft tissues such as liver, kidney, brain and spleen [5], for this reason we select the liver and kidney to describe the histological changes after lead exposure. The

liver via the portal vein is the first organ exposed to externally absorbed nutrients and other xenobiotics. The liver is composed of highly active metabolic tissue containing huge complement of detoxification machinery system [12].

The most common findings in liver were vacuolation, fatty degenerative, congestion within central veins and a losing normal architecture into the hepatocytes. These findings were in agreement with other researchers [11, 13, 14], these changes may be because of that lead acetate was exhibited to decline cytochrome P450 content [12]. Also lead induces mitogenic response in the rodent liver [15]. Lead acetate was found to induce glutathione-S-transferase in rat liver [16].

Histopathological investigations of liver display changes which reflect damages in hepatic tissues possibly due to cycling of heavy metal and also observed the same histological changes in liver in rat after exposure to heavy metals. It has been presented that heavy metals forms mercaptides with the SH groups of cysteine and less stable complexes with other amino acid chain and these changes reflect damages in hepatic tissues [17].

### References

- [1] Hurst, H. E., Martin, M. D. 2004. Toxicology in Yagiela, J. A. ; Dowd, F. I. Neidle, E. A. Pharmacology and Therapeutic for Dentistry. 5th Edn., Mosby, USA., pp. 829-48.
- [2] Silbergeld, E. K. 2005. Learners and learning in the twenty-first century: what do we know about students' attitudes towards and experiences of information and communication technologies that will help us design courses? *Stud High Educ*, 30(3): 257-74.
- [3] Riaz, F., Khan, U. A., Ayub, M., Shaukat, S. 2011. Protective role of ginger on lead induced derangement in plasma testosterone and luteinizing hormone levels of male Sprague Dawley rats. *J Ayub Med Coll Abbottabad*, 23(4).
- [4] Tian, L., Lawrence, D. A. 1995. Lead inhibits nitric oxide production in vitro by murine splenic macrophages. *ToxicolApplPharmacol*, 132(1): 156-63.
- [5] Kosnett, M. J. 2004. Heavy Metal Intoxication and Chelators. In Katzung, B. G. Basic and Clinical Pharmacology. Mc Graw-Hill, New York. pp. 970-81.
- [6] Meyer, P. A., Brown, M. J., Falk, H. 2008. Global approach to reducing lead exposure and poisoning. *Mutat Res - Rev Mut Res*, 659(1): 166-75.
- [7] Goyer, R. A., Clarkson, T. W. 1996. Toxic effects of metals. Casarett&Doull's Toxicology The Basic Science of Poisons, Fifth Edition, Klaassen, CD [Ed] McGraw-Hill Health Professions Division, ISBN, 71054766: pp. 811-67.
- [8] Loghman-Adham, M. 1997. Renal effects of environmental and occupational lead exposure. *Environmental Health Perspectives*, 105(9): 923-8.
- [9] Gidlow, D. A. 2004. Lead Toxicity. *Depth Review Occupational Medicine*, 54: 76-81.
- [10] Plumlee, K. H. 2004. Metals and Minerals in Clinical Veterinary Toxicology, 1st Edn. Mosby, U.S.A. pp. 193-230.

- [11] Durgut, R., Koc, A., Gonenci, R., Bal, R., Celik, S., Guzel, M., et al. 2008. Effects of high dose lead toxication on liver, kidneys, heart, brain and blood in rabbits: an experimental study. *Journal of Applied Biological Sciences*, 2(2): 11-8.
- [12] US-EPA. 1986. Air quality criteria document for lead (Pb), Vol. 4. Washington DC: US Environmental Protection Agency. PP. 264-7.
- [13] Piasek, M., Kostial, K., Bunarević, A. 1989. The effect of lead exposure on pathohistological changes in the liver and kidney in relation to age in rats. *Arhivzahigijjenuradaitoksikologiju*, 40(1): 15-21.
- [14] Kojima, M., Sekikawa, K., Nemoto, K., Degawa, M. 2005. Tumor necrosis factor- $\alpha$ -independent downregulation of hepatic cholesterol 7 $\alpha$ -hydroxylase gene in mice treated with lead nitrate. *Toxicological Sciences*, 87(2): 537-42.
- [15] Calabrese, E. J., Baldwin, L. A. 1992. Lead-induced cell proliferation and organ-specific tumorigenicity. *Drug Metabolism Reviews*, 24(3): 409-16.
- [16] Suzuki, T., Morimura, S., Diccianni, M. B., Yamada, R., Hochi, S.-i., Hirabayashi, M., et al. 1996. Activation of glutathione transferase P gene by lead requires glutathione transferase P enhancer I. *Journal of Biological Chemistry*, 271(3): 1626-32.
- [17] Gajawat, S., Sancheti, G., Goyal, P. 2005. Vitamin; C against Concomitant Exposure to Heavy Metal and Radiation: A Study on Variations in Hepatic Cellular Counts. *Asian Journal of Experimental Sciences*, 19(2): 53-8.